

Review

Bioactive compounds in banana fruits and their health benefits

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Abstract

Banana is an edible fruit and is herbaceous flowering plant belonging to the genus *Musa* and the family *Musaceae*. Banana is also eaten as cooked vegetable (and is then called plantains). All the edible banana fruits are seedless (parthenocarpic) and belong to two main species, *Musa acuminata* Colla and *Musa balbisiana* Colla. The hybrid from these two species *Musa x paradisiaca* L. is also available nowadays. Although banana is native to Indomalaya and Australia, Papua New Guinea was the first to domesticate this fruit. Banana has now spread to almost 135 countries around the world. As per 2016 data, nearly 28 per cent of the total world's banana production comes from India and China. Cavendish group banana, being the main export item from the banana-exporting countries, usually refers to soft, sweet, and dessert banana in the Western countries, but the plantain bananas have firm, starchy fruit which is suitable for cooking as a vegetable. Banana is known to be rich not only in carbohydrates, dietary fibres, certain vitamins and minerals, but is also rich in many health-promoting bioactive phytochemicals. General composition including various bioactives and their health contributions has been reviewed in this paper.

Key words: banana fruit; nutritional value; phytochemicals; bioactives; phenolics; biogenic amines.

Introduction

The consumption of fruits and fruit products is known not only to promote general good health but also lower the risk of various chronic diseases, such as heart diseases, stroke, gastrointestinal disorders, certain types of cancer, hypertension, age-related macular degeneration, cataract of the eye, skin conditions, lowering of low-density lipoprotein (LDL) cholesterol, and improved immune function. To promote healthy eating lifestyle, the USDA recommends filling up half the plate with fruits and vegetables, because these provide a good amount of dietary fibres, certain vitamins (*e.g.* ascorbic acid, folic acid, and vitamin A precursors), many minerals (*e.g.* potassium, magnesium, iron, and calcium), and many other important phytochemicals having strong antioxidative properties. Fruits make an important part of the balanced diet adopted by the humans. USDA recommends daily five servings of fruits to obtain most of the health benefits. Depending upon their origin and production area temperature, fruits are classified into temperate fruits, sub-tropical fruits, and tropical fruits. Banana belongs to the tropical fruits as it grows more profusely in tropical rain forest areas.

Interestingly, banana fruit has flesh not only rich in starch which changes into sugars on ripening but is also a good source of resistant starch. Banana is known to be rich in carbohydrates, dietary fibres, certain vitamins, and minerals (Table 1). The presence of various bioactive phytochemicals and their nutritional significance has been discussed in this review paper (Figure 1).

Antioxidant Compounds in Banana Fruits

The reactive oxygen species (ROS) and reactive nitrogen species (RON), such as hydroxyl radicals, superoxide ions, nitric oxide radicals, and singlet oxygen and hydrogen peroxide, have now been implicated in the causation of many disorders like arthritis, diabetes, arteriosclerosis, age-related macular degeneration, certain types of cancer, inflammation, genotoxicity, and Alzheimer disease. The exact mechanism is not known but the reaction of these ROS and RNS species with biomolecules such as lipids, proteins, and DNA may be the cause of these disease conditions (Shukla *et al.*, 2009; Septembre-Malaterre *et al.*, 2016). Kandaswamy and Aradhya (2014) have

Table 1. Chemical composition of banana fruit (as is basis per 100 g)

Constituent	Amount µg, mg, g, or percent daily value
Energy	371 kJ (89 kcal)
Water	74.91 g
Carbohydrates	22.84 g
Sugars	12.23 g
Dietary fibre	2.6 g
Vitamins	
Pantothenic acid (B5)	0.334 mg, (7%)
Pyridoxine (B6)	0.4 mg, (31%)
Choline	9.8 mg, (2%)
Vitamin C	8.7 mg, (10%)
Minerals	
Magnesium	27 mg, (8%)
Phosphorus	22 mg, (3%)
Potassium	358 mg, (8%)
Sodium	1 mg, (0%)
Zinc	0.15 mg, (2%)

Adopted from: Wikipedia, Internet, USDA databases.

shown the banana rhizome to be a rich source of many polyphenolic compounds having antioxidant activities. Pazmino-Duran *et al.* (2001) have suggested the use of anthocyanins from banana bracts (florets) as natural colourants. They identified various anthocyanins such as cyanidin-3-rutinoside (main one as 80 per cent of total pigments, being 32.3 mg/100 g) and 3-rutinoside derivatives of delphinidin, pelargonidin, peonidin, and malvidin. Interestingly, the addition of heat-treated onion extract was found to inhibit the polyphenol oxidase (PPO) during ripening of banana fruit at room temperature (Lee, 2007). Even the Maillard reaction products (MRP) significantly affected the banana PPO activity. The phytochemistry and pharmacology of wild banana (*Musa acuminata* Colla) have been reviewed by Mathew and Negi (2017) and they suggested the use of banana pulp and peel for the development of drugs and use in functional foods.

Not only banana pulp, but pseudo stem and banana fruit peel have been found to be the good sources of antioxidants (Table 2). Aziz *et al.* (2011) have analysed the native banana pseudo-stem flour (NBPF) and tender core of pseudo-stem flour (TCBPF) for chemical and functional properties. They found higher content of polyphenols, flavonoids, total dietary fibre, insoluble dietary fibre, lignin, hemicellulose, cellulose, antioxidant capacity, and free-radical scavenging



Figure 1. Banana tree and banana fruits of various maturities. (Source: Internet Wikipedia.)

capacity in NBPF than TCBPF. In exhaustive reviews, [Pereira and Maraschin \(2015\)](#) and [Singh et al. \(2016\)](#) have reported that banana is rich in many bioactive compounds, such as carotenoids, flavonoids, phenolics, amines, vitamin C, and vitamin E having antioxidant activities to provide many human health benefits. Recently, [Vu et al. \(2018\)](#) have also reviewed the phenolic compounds and their potential health benefits coming from banana peel. They have suggested the use of this valuable by-product from banana fruit processing industry in food and pharmaceutical industry. [Anyasi et al. \(2018\)](#) have analysed the essential macro and trace minerals as well as phenolic compounds in unripe banana flour obtained from the pulp of four cultivars treated with ascorbic, citric, and lactic acids before drying in a forced air dryer at 70°C. Results of their liquid chromatography-mass spectrometry-electrospray ionization (LC-MS-ESI) assay of phenolics revealed the presence of two flavonoids, epicatechin and 3-O-rhamnosyl-glucoside in varying concentrations. Among the essential minerals, zinc had the lowest concentration of 3.55 mg/kg, but the potassium was the highest, 14746.73 mg/kg in these cultivars.

Carotenoids

Carotenoids is a class of compounds having some 600 members in this family. Some of these are precursors for vitamin A, and others are known to have strong antioxidant capacity to scavenge ROS. Among the carotenoids present in banana fruit, α -carotene, β -carotene, and β -cryptoxanthin have provitamin A activity, but others like lycopene and lutein have a strong antioxidant capacity ([Erdman et al., 1993](#)). Lycopene is known to provide protection against prostate cancer among men, and lutein offers human health benefits to serve as an inhibitor of age-related macular degeneration ([Davey et al., 2006](#)). Later, [Davey](#)

[et al. \(2009\)](#) have analysed 171 different genotypes of *Musa* spp. for provitamin A carotenoids and 47 genotypes for two minerals (iron and zinc). They found a great variability in provitamin A among the various cultivars, but a low variability in iron and zinc, irrespective of the soil type and growing environmental conditions. They suggested the use of high provitamin A and trace mineral cultivars as development strategies to improve the nutritional health and alleviation of micronutrient deficiencies among the *Musa*-consuming populations.

Yellow- and orange-fleshed banana cultivars are known to be richer in trans- β -carotene content ([Englberger et al., 2006](#)). Carotenoid content of some of the banana cultivars is presented in [Table 3](#). Consumption of fruits rich in carotenoids is reported to boost immunity and reduce the risk of various diseases, such as cancer, type II diabetes, and cardiovascular problems ([Krinsky and Johnson, 2005](#)). Certain banana cultivars rich in provitamin A carotenoids can be grown and consumed by the poor population of the world that is having serious vitamin A deficiency, and the consumption of such banana fruit would alleviate vitamin A deficiency ([Fungo and Pillay, 2013](#)).

Phenolic Compounds

Phenolics present in banana fruit are the major bioactive compounds having antioxidant properties and are known for providing health benefits ([Table 4](#)). Various phenolics present in banana have been identified as follows: gallic acid, catechin, epicatechin, tannins, and anthocyanins. Banana rhizome is used as food and for medicinal properties as well in South India as it is very rich in phenolics ([Kandasamy and Aradhya, 2014](#)). [Russel et al. \(2009\)](#) have detected many phenolics in banana, such as ferulic, sinapic, salicylic, gallic, p-hydroxybenzoic, vanillic, syringic, gentisic, and p-coumaric acids as major components.

Table 2. Antioxidant activity, total polyphenol, and individual polyphenolic compounds present in organic acid treated (20 g/l) unripe banana flour

Antioxidant and phenolic content	Cultivar	Ascorbic acid	Citric acid	Lactic acid
DPPH (mg GA/g DW)	Luvhele	0.84 ^c ± 0.01	0.90 ^d ± 0.04	0.95 ^e ± 0.01
	Mabonde	0.81 ^c ± 0.00	0.89 ^f ± 0.01	0.96 ^g ± 0.02
Total polyphenols (mg GAE/100g DW)	Luvhele	777.83 ^{de} ± 56.41	707.87 ^c ± 12.62	769.24 ^{cd} ± 18.68
	Mabonde	1081.54 ^b ± 23.18	856.46 ^f ± 18.01	929.36 ^g ± 9.32
Epicatechin (µg/g DW)	Luvhele	3.49 ^a ± 0.67	4.05 ^a ± 0.76	4.24 ^a ± 0.84
	Mabonde	1.35 ^a ± 0.18	–	–
Myricetin-3-O-rhamnosyl glucoside (µg/g DW)	Luvhele	17.08 ^a ± 1.97	13.91 ^{bc} ± 1.61	9.91 ^b ± 1.58
	Mabonde	13.69 ^a ± 1.61	5.68 ^{bc} ± 1.04	8.20 ^b ± 0.96

Means with different letters across rows are significantly different at $P < 0.05$. Values are Means ± SE of triplicate measurements. DPPH, 1,1-diphenyl-2-picrylhydrazyl. (Adopted and modified from [Anyasi et al., 2018](#).)

Table 3. Carotenoid content of different banana cultivars (µg/100 g)

Cultivar	Trans- α carotene	Trans- β carotene	Total carotenoid content	Reference	
Hung Tu ripe pulp	1849 FW	5653 FW	7760 FW	Beatrice et al., 2015	
To'o ripe pulp	2055 FW	5267 FW	7765 FW		
Sepi ripe pulp	4728 FW	5611 FW	10,067 FW	Englberger et al., 2010	
Apantu ripe pulp	3287 FW	6387 FW	10,056 FW		
Bungaoisan ripe pulp	779 FW	857 FW	1675 FW		
Aibwo	2358 FW	5945 FW	9400 FW		
Fagufagu	1524 FW	3428 FW	5054 FW		
Ropa	3682 FW	1324 FW	5218 FW		
Gatagata	79 FW	695 FW	774 FW		
Toraka Parao	250 FW	526 FW	776 FW		
Red banana peel and pulp	ND	123 and 29.6 DW	200 and 250 DW		Arora et al., 2008

Source: Adopted and modified from: [Singh et al., 2016](#).

However, ferulic acid content was the highest (69 per cent of cinnamic acids) among these phenolics. Banana peel is also a rich source of phenolic compounds. Tsamo *et al.* (2015) analysed banana pulp and peel from nine plantain cultivars and two dessert banana cultivars. According to their results, hydroxycinnamic derivatives, such as ferulic acid-hexoside, were the major ones (4.4–85.1 µg/g DW) in plantain pulp. They observed large variations in the phenolic contents among the cultivars tested. In the peel from plantain cultivars, rutin was the most abundant flavonol glycoside (242.2–618.7 µg/g DW). Thus, the banana peel and pulp both are good sources of health-promoting phenolic compounds. Among the flavonoids detected in banana are as follows: quercetin, myricetin, kaempferol, and cyanidin which provide health benefits mainly because they act as free radicals, ROS, and RNS scavengers (Kevers *et al.*, 2007). Most of these phenolics are known to also exhibit antibacterial, antiviral, anti-inflammatory, antiallergenic, antithrombotic, and vasodilatory activities (Cook and Sammon, 1996). Sulaiman *et al.* (2011) have determined the total phenolic and mineral contents in pulp and peel from eight banana (*Musa* spp.) cultivars grown in Malaysia. With a few exceptions, the peel extracts had the higher total phenolics and total antioxidant activities than the pulp. Among minerals, potassium was the major element found in both the peel and pulp followed by phosphorus, magnesium, and sodium.

Health Benefits of Bioactive Components in Banana Fruits

Health benefits of phenolics

A flavonoid, leucocyanidin, has been identified as a predominant component of aqueous extract of unripe banana pulp that showed significant anti-ulcerogenic activity (Lewis *et al.*, 1999). Thus, many flavonoids, especially leucocyanidin analogues, may offer immense therapeutic potential in the treatment of gastric disease conditions.

The structure–activity relationship of flavonoids indicates that their antioxidant capacity, scavenging free radicals, and chelating action are related to the presence of functional groups in their nuclear structure (Heim *et al.*, 2002). They also attributed most of the health

benefits from the consumption of flavonoids to their antioxidant and chelating properties. Because of these properties, flavonoids are also shown to exhibit antimutagenic and antitumoral activities (Rice-Evans *et al.*, 1996). The flavonoids can also inhibit many enzymes, such as oxygenases (prostaglandin synthase), required in the synthesis of eicosanoids. Thus, the flavonoids inhibit hyaluronidase activity and help in maintaining the proteoglycans of connective tissues. This would prevent the spread of bacterial or tumour metastases (Havsteen, 2002). As the flavonoids get preferentially oxidized, they are reported to prevent the oxidation of body's natural water-soluble antioxidants like ascorbic acid (Korkina and Afanas'ev, 1997). Generally, after the consumption of banana fruit, the peel ends up as a feed for the animals only. The disposal of peel (pomace) and other by-products from banana-processing industry causes a serious environmental problem (Zhang *et al.*, 2005). Banana peel is reported to be rich in many high-value health-promoting antioxidant phytochemicals, such as anthocyanins, delphinidin, and cyanidins (Seymour, 1993). In a recent study, Rebello *et al.* (2014) have also shown the banana peel extract to be a rich source of total phenolics (29 mg/g as GAE), which are responsible for the very high antioxidant activity. They also determined various other antioxidant compounds, namely, highly polymerized prodelphinidins (~3952 mg/kg), flavonol glycosides (mainly 3-rutinosides and predominantly quercetin-based compounds, ~129 mg/kg), B-type pro-cyanidin dimers, and monomeric flavan-3-ols (~126 mg/kg).

Health benefits of biogenic amines

Banana peel and pulp are known to be good sources of certain biogenic amines (catecholamines) which are produced by the decarboxylation of amino acids or by the amination of aldehydes and ketones. Catecholamines include dopamine, serotonin, epinephrine, and norepinephrine and are reported to occur in many plants in considerable amounts (Ponchet *et al.*, 1982). In animals, these biogenic amines are reported to work as neurotransmitters for the hormonal regulation of glycogen metabolism (Kimura, 1968). When banana is consumed by humans, serotonin present in the pulp (ranging from 8 to 50 µg/g)

Table 4. Uses and health benefits of bioactive compounds in banana

Bioactive compound	Health benefits	Reference source
Tannic acid	Applied as medicinal agents for the treatment of burns	Siang (1983)
Catechin	Resistance of LDL to oxidation, brachial artery dilation increased plasma anti-oxidant activity, and fat oxidation	Williamson and Manach (2005)
Gallic acid	Antioxidant and potential hepatoprotective effects	Rasool <i>et al.</i> (2010)
Cinnamic acid	Is a precursor to the sweetener aspartame by the means of enzyme catalysed amination to phenylalanine	Garbe (2000)
p-Coumaric acid	Antioxidant properties and potentially reduce the risk of stomach cancer	Ferguson, Zhu, and Harris (2005)
Gallocatechin gallate	Cholesterol reduction	Ikeda <i>et al.</i> (2003)
Quercetin	Promotes overall cardiovascular health by encouraging blood flow	Perez-Vizcaino and Duarte (2010)
Ferulic acid	Antioxidant, antimicrobial, anti-inflammatory, antiallergic, anticarcinogenic, modulation of enzyme activity, antiviral and vasodilatory actions	Kumar and Pruthi (2014)
Trans- α carotene	Precursor to vitamin A	Li <i>et al.</i> (2011)
Trans- β carotene	Reduce the risk of CVD and cancer	Li <i>et al.</i> (2011)
Violaxanthin	Used as a food colourant	
Cryptoxanthin	Food colourant might reduce the risk of lung cancer	DeLorenze <i>et al.</i> (2010)
Serotonin	Might contribute to feelings of well-being and happiness	Young (2007)
Dopamine	Reduce the plasma oxidative stress and enhance the resistance to oxidative modification of LDL	Yin <i>et al.</i> , (2008)
Catecholamines	Increases blood pressure, glucose levels, and heart beat rate	Kuklin and Conger (1995)
β -Sitosterol	Potential to reduce blood cholesterol levels and benign prostatic hyperplasia (BPH)	Wilt <i>et al.</i> (1999)
Campesterol and stigmasterol	Reduces the absorption of cholesterol in the human intestines	Choudhary and Tran (2011)

creates a feeling of well-being and happiness. Banana contains a large amount of dopamine and norepinephrine (Buckley, 1961). Waalkes *et al.* (1958) were the first to report the amount of various catecholamines in banana pulp as follows: serotonin, 28 µg/g; norepinephrine, 1.9 µg/g; and dopamine, 7.9 µg/g. The concentrations of dopamine in the pulp of yellow banana (*M. acuminata*), red banana (*Musa sapientum*), and plantain has been reported to be 42, 54, and 5.5 µg/g, respectively (Feldman *et al.*, 1987). They highlighted the role of dopamine in human brain and body as a neurotransmitter having a strong influence on mood and emotional stability. Dopamine in the peel and pulp of commercially ripened *Musa Cavendish* is reported to range from 80 to 560 mg/100 g, and 2.5 to 10 mg/100 g, respectively (Kanazawa and Sakakibara, 2000). Tryptophan being one of the precursors for the synthesis of dopamine, the presence of this amino acid in banana peel increases the interest in possibilities of preventing neurodegenerative diseases like Parkinson's using this by-product of food-processing industry by developing pharmaceutical formulations. However, the increase in dopamine content from unripe to the ripened stage in both the peel and pulp has been reported by many workers (Romphophak *et al.*, 2005; Gonzalez-Montelongo *et al.*, 2010). They also suggested that the decline in dopamine concentration during over-ripening stage may be due to its oxidation to quinones which may further polymerize to melanin pigments.

Using peroxide value and thiobarbituric activity determination, the antioxidant compounds present in water extract of banana peel have shown to suppress the autooxidation of linoleic acid by 65 to 70 per cent after 5 days of incubation (Kanazawa and Sakakibara, 2000). When they compared dopamine with other natural antioxidants, such as ascorbic acid, reduced glutathione, and phenolic acids (e.g. gallo-catechin gallate), the dopamine showed higher antioxidant activity *in vitro* (DPPH assay). Gonzalez-Montelongo *et al.* (2010) have reported the banana peel extracts to be rich in dopamine, L-dopa, and catecholamines with a significant antioxidant capacity. They found no significant difference in the antioxidant activity in the banana peel extracts from different cultivars. The biogenic amines are also shown to play an important role in offering plants' resistance to various invading pathogens through their interaction with phytohormones (*via* auxin oxidation), thus affecting the growth and development of plants (Newman *et al.*, 2001; Roepenack-Lahaye *et al.*, 2003).

Health benefits of phytosterols

These naturally occurring plant sterols have attracted the attention of food manufacturers to produce functional foods having higher health benefits. Because of their structural similarity with cholesterol, they compete with cholesterol for absorption in the gut, thus lowering the blood cholesterol levels (Marangoni and Poli, 2010). They reported that a daily intake of 3 g of phytosterols results in marked reduction of LDL cholesterol levels. Various phytosterols reported in the banana peel are stigmasterol, -sitosterol, campesterol, 24-methylene cycloartenol, cyclooleucenol, and cycloartenol (Knapp and Nicholas, 1969). Now the health professionals recommend the consumption of plant sterols-rich diet to lower the LDL cholesterol in patients who do not tolerate cholesterol-lowering statin drugs (Ostlund *et al.*, 2003). Banana fruit has been shown to contain a good amount of phytosterols both in the peel and pulp (Akihisa *et al.*, 1986). The phytosterols content in unripe banana in the range of 2.8 to 12.4 g/kg DW has been reported by Vilaverde *et al.* (2013). According to their results, the *Musa balbisiana* cultivars, such as 'Dwarf Red' and 'Silver', had higher amounts of phytosterols than the *M. acuminata* cultivars. The lipophilic extract of ripe banana pulp from several cultivars of the *M. acuminata* and *M. balbisiana* species has been found

to be a source of ω-3 and ω-6 fatty acids, phytosterols, long-chain aliphatic alcohols, and α-tocopherol, thus offering well-established nutritional and health benefits (Vilela *et al.*, 2014).

Summary

The above discussion brings out the importance of consuming banana fruits for obtaining various health benefits. It is not only the banana fruit pulp, but also the peel of this fruit is known to contain many important phytochemicals and offers many health benefits. More research is needed to be carried out to find ways of using banana fruit peel in the development of many new functional foods.

References

- Akihisa, T., Shimizu, N., Tamura, T., Matsumoto, T. (1986). (24S)-14a, 24-Dimethyl-9b, 19-cyclo-5a-cholest-25-en-3b-ol: a new sterol and other sterols in *Musa sapientum*. *Lipids*, 21: 494–497.
- Anyasi, T. A., Jideani, A. I. O., Mchau, G. R. A. (2018). Phenolics and essential mineral profile of organic acid pretreated unripe banana flour. *Food Research International (Ottawa, Ont.)*, 104: 100–109.
- Arora, A., Choudhary, D., Agarwal, G., Singh, V. P. (2008). Compositional variation in β-carotene content, carbohydrate and antioxidant enzymes in selected banana cultivars. *International Journal of Food Science and Technology*, 43: 1913–1921.
- Aziz, N. A. A., Ho, L. H., Azahari, B., Bhat, R., Cheng, L. H., Ibrahim, M. N. M. (2011). Chemical and functional properties of the native banana (*Musa acuminata* × *balbisiana* Colla cv. Awak) pseudo-stem and pseudo-stem tender core flours. *Food Chemistry*, 128: 748–753.
- Beatrice, E., Deborah, N., Guy, B. (2015). Provitamin A carotenoid content of unripe and ripe banana cultivars for potential adoption in eastern. *African Journal of Food Composition and Analysis*, 43: 1–6.
- Buckley, E. H. (1961). Further studies on the biosynthesis of 3-hydroxytyramine in the peel of the banana. *Plant Physiology*, 36: 315–320.
- Choudhary, S. P., Tran, L. S. (2011). Phytosterols: perspectives in human nutrition and clinical therapy. *Current Medicinal Chemistry*, 18: 4557–4567.
- Cook, N. C., Sammon, S. (1996). Flavanoids chemistry, metabolism, cardioprotective effects, and dietary sources. *Nutritional Biochemistry*, 7: 66–76.
- Davey, M. W., Bergh, I. V., Markham, R., Swennen, R., Keulemans, J. (2009). Genetic variability in *Musa* fruit provitamin A carotenoids, lutein and mineral micronutrient contents. *Food Chemistry*, 115: 806–813.
- Davey, M. W., Keulemans, J., Swennen, R. (2006). Methods for the efficient quantification of fruit provitamin A contents. *Journal of Chromatography A*, 1136: 176–184.
- DeLorenze, G. N., *et al.* (2010). Daily intake of antioxidants in relation to survival among adult patients diagnosed with malignant glioma. *BMC Cancer*, 10: 215.
- Englberger, L., Wills, R. B., Blades, B., Dufficy, L., Daniells, J. W., Coyne, T. (2006). Carotenoid content and flesh color of selected banana cultivars growing in Australia. *Food and Nutrition Bulletin*, 27: 281–291.
- Erdman, J. W. Jr, Bierer, T. L., Gugger, E. T. (1993). Absorption and transport of carotenoids. *Annals of the New York Academy of Sciences*, 691: 76–85.
- Feldman, J. M., Lee, E. M., Castleberry, C. A. (1987). Catecholamine and serotonin content of foods: effect on urinary excretion of homovanillic and 5-hydroxyindoleacetic acid. *Journal of the American Dietetic Association*, 87: 1031–1035.
- Ferguson, L. R., Zhu, S. T., Harris, P. J. (2005). Antioxidant and antigenotoxic effects of plant cell wall hydroxycinnamic acids in cultured HT-29 cells. *Molecular Nutrition & Food Research*, 49: 585–593.
- Fungo, R., Pillay, M. (2013). β-Carotene content of selected banana genotypes from Uganda. *African Journal of Biotechnology*, 10: 5423–5430.
- Garbe, D. (2000). Cinnamic Acid. *Ullmann's Encyclopedia of Industrial Chemistry*. Wiley-VCH Verlag GmbH & Co, KGaA.
- González-Montelongo, R., Lobo, M. G., González, M. (2010). Antioxidant activity in banana peel extracts: testing extraction conditions and related bioactive compounds. *Food Chemistry*, 119: 1030–1039.

- Havsteen, B. H. (2002). The biochemistry and medical significance of the flavonoids. *Pharmacology & Therapeutics*, 96: 67–202.
- Heim, K. E., Tagliaferro, A. R., Bobilya, D. J. (2002). Flavonoid antioxidants: chemistry, metabolism and structure-activity relationships. *The Journal of Nutritional Biochemistry*, 13: 572–584.
- Ikedo, I., et al. (2003). Heat-epimerized tea catechins rich in gallic acid gallate and catechin gallate are more effective to inhibit cholesterol absorption than tea catechins rich in epigallocatechin gallate and epicatechin gallate. *Journal of Agricultural and Food Chemistry*, 51: 7303–7307.
- Kanazawa, K., Sakakibara, H. (2000). High content of dopamine, a strong antioxidant, in Cavendish banana. *Journal of Agricultural and Food Chemistry*, 48: 844–848.
- Kandasamy, S., Aradhya, S. M. (2014). Polyphenolic profile and antioxidant properties of rhizome of commercial banana cultivars grown in India. *Food Bioscience*, 8: 22–32.
- Kevers, C., Falkowski, M., Tabart, J., Defraigne, J. O., Dommès, J., Pincemail, J. (2007). Evolution of antioxidant capacity during storage of selected fruits and vegetables. *Journal of Agricultural and Food Chemistry*, 55: 8596–8603.
- Kimura, M. (1968). Fluorescence histochemical study on serotonin and catecholamine in some plants. *Japanese Journal of Pharmacology*, 18: 162–168.
- Knapp, F. F., Nicholas, H. J. (1969). The sterols and triterpenes of banana peel. *Phytochemistry*, 8: 207–214.
- Korkina, L. G., Afanas'ev, I. B. (1997). Antioxidant and chelating properties of flavonoids. *Advances in Pharmacology (San Diego, Calif.)*, 38: 151–163.
- Krinsky, N. I., Johnson, E. J. (2005). Carotenoid actions and their relation to health and disease. *Molecular Aspects of Medicine*, 26: 459–516.
- Kuklin, A. I., Conger, B. V. (1995). Catecholamines in plants. *Journal of the Plant Growth Regulation*, 14: 91–97.
- Kumar, N., Pruthi, V. (2014). Potential applications of ferulic acid from natural sources. *Biotechnology Reports (Amsterdam, Netherlands)*, 4: 86–93.
- Lee, M. K. (2007). Inhibitory effect of banana polyphenol oxidase during ripening of banana by onion extract and Maillard reaction products. *Food Chemistry*, 102: 146–149.
- Lewis, D. A., Fields, W. N., Shaw, G. P. (1999). A natural flavonoid present in unripe plantain banana pulp (*Musa sapientum* L. Var. *paradisica*) protects the gastric mucosa from aspirin-induced erosions. *Journal of Ethnopharmacology*, 65: 283–288.
- Li, C., Ford, E. S., Zhao, G., Balluz, L. S., Giles, W. H., Liu, S. (2011). Serum α -carotene concentrations and risk of death among US adults: the third national health and nutrition examination survey follow-up study. *Archives of Internal Medicine*, 171: 507–515.
- Marangoni, F., Poli, A. (2010). Phytosterols and cardiovascular health. *Pharmacological Research*, 61: 193–199.
- Mathew, N. S., Negi, P. S. (2017). Traditional uses, phytochemistry and pharmacology of wild banana (*Musa acuminata* Colla): a review. *Journal of Ethnopharmacology*, 196: 124–140.
- Newman, M. A., von Roepenack-Lahaye, E., Parr, A., Daniels, M. J., Dow, J. M. (2001). Induction of hydroxycinnamoyl-tyramine conjugates in pepper by *xanthomonas campestris*, a plant defense response activated by hrp gene-dependent and hrp gene-independent mechanisms. *Molecular Plant-Microbe Interactions*, 14: 785–792.
- Oliveira, T. Í., et al. (2016). Optimization of pectin extraction from banana peels with citric acid by using response surface methodology. *Food Chemistry*, 198: 113–118.
- Ostlund, R. E. Jr, Racette, S. B., Stenson, W. F. (2003). Inhibition of cholesterol absorption by phytosterol-replete wheat germ compared with phytosterol-depleted wheat germ. *The American Journal of Clinical Nutrition*, 77: 1385–1389.
- Pazmino-Duran, E. A., Giusti, M. M., Wrolstad, R. E., Gloria, M. B. A. (2001). Anthocyanins from banana bracts (*Musa x paradisica*) as potential food colorants. *Food Chemistry*, 73: 327–332.
- Pereira, A., Maraschin, M. (2015). Banana (*Musa* spp) from peel to pulp: ethnopharmacology, source of bioactive compounds and its relevance for human health. *Journal of Ethnopharmacology*, 160: 149–163.
- Perez-Vizcaino, F., Duarte, J. (2010). Flavonols and cardiovascular disease. *Molecular Aspects of Medicine*, 31: 478–494.
- Ponchet, M., Martin-Tanguy, J., Marais, A., Martin, C. (1982). Hydroxycinnamoyl acid amides and aromatic amines in the inflorescences of some *Araceae* species. *Phytochemistry*, 21: 2865–2869.
- Rasool, M. K., et al. (2010). Hepatoprotective and antioxidant effects of gallic acid in paracetamol-induced liver damage in mice. *The Journal of Pharmacy and Pharmacology*, 62: 638–643.
- Rebello, L. P. G., Ramos, A. M., Pertuzatti, P. B., Barcia, M. T., Castillo-Munoz, N., Hermosin-Gutierrez, I. (2014). Flour of banana (*Musa AAA*) peel as a source of antioxidant phenolic compounds. *Food Research International*, 55: 397–403.
- Rice-Evans, C. A., Miller, N. J., Paganga, G. (1996). Structure-antioxidant activity relationships of flavonoids and phenolic acids. *Free Radical Biology & Medicine*, 20: 933–956.
- Romphopha, T., Siriphanich, J., Ueda, Y., Abe, K., Chachin, K. (2005). Changes in concentrations of phenolic compounds and polyphenol oxidase activity in banana peel during storage. *Food Preservation Science*, 31: 111–115.
- Russell, W. R., Labat, A., Scobbie, L., Duncan, G. J., Duthie, G. G. (2009). Phenolic acid content of fruits commonly consumed and locally produced in Scotland. *Food Chemistry*, 115: 100–104.
- Von Roepenack-Lahaye, E., et al. (2003). P-coumaroylnoradrenaline, a novel plant metabolite implicated in tomato defense against pathogens. *The Journal of Biological Chemistry*, 278: 43373–43383.
- Septembre-Malaterre, A., Stanislas, G., Douraguia, E., Gonthier, M. P. (2016). Evaluation of nutritional and antioxidant properties of the tropical fruits banana, litchi, mango, papaya, passion fruit and pineapple cultivated in réunion french island. *Food Chemistry*, 212: 225–233.
- Seymour, G. B. (1993). Banana. In: Seymour, J. E., Tucker, G. A. (eds.) *Biochemistry of Fruit Ripening*. Chapman and Hall, NY, pp. 83–106.
- Shukla, S., Mehta, A., John, J., Singh, S., Mehta, P., Vyas, S. P. (2009). Antioxidant activity and total phenolic content of ethanolic extract of *caesalpinia bonducella* seeds. *Food and Chemical Toxicology*, 47: 1848–1851.
- Siang, S. T. (1983). Use of combined traditional Chinese and Western medicine in the management of burns. *Panminerva medica*, 25: 197–202.
- Singh, B., Singh, J. P., Kaur, A., Singh, N. (2016). Bioactive compounds in banana and their associated health benefits – A review. *Food Chemistry*, 206: 1–11.
- Sulaiman, S. F., Yusoff, N. A. M., Eldeen, I. M., Seow, E. M., Sajak, A. A. B., Supriatno, Ooi, K. L. (2011). Correlation between total phenolic and mineral contents with antioxidant activity of eight Malaysian bananas (*Musa* sp.). *Journal of Food Composition and Analysis*, 24: 1–10.
- Passo Tsamo, C. V., et al. (2015). Phenolic profiling in the pulp and peel of nine plantain cultivars (*Musa* sp.). *Food Chemistry*, 167: 197–204.
- Vilaverde, J. J., et al. (2013). High valuable compounds from the unripe peel of several *Musa* species cultivated in Madeira Island (Portugal). *Industrial Crops Production*, 42: 507–512.
- Vilela, C., et al. (2014). Lipophilic phytochemicals from banana fruits of several *Musa* species. *Food Chemistry*, 162: 247–252.
- Vu, H. T., Scarlett, C. J., Vuong, Q. V. (2018). Phenolic compounds within banana peel and their potential uses: a review. *Journal of Functional Foods*, 40: 238–248.
- Waalkes, T. P., Sjoerdsma, A., Creveling, C. R., Weissbach, H., Udenfriend, S. (1958). Serotonin, norepinephrine, and related compounds in bananas. *Science (New York, N.Y.)*, 127: 648–650.
- Williamson, G., Manach, C. (2005). Bioavailability and bioefficacy of polyphenols in humans. II. Review of 93 intervention studies. *The American Journal of Clinical Nutrition*, 81: 243S–255S.
- Wilt, T., Ishani, A., MacDonald, R., Stark, G., Mulrow, C., Lau, J. (1999). Betasitosterols for benign prostatic hyperplasia. *Cochrane Database Systematic Reviews*, 3.
- Yin, X., Quan, J., Kanazawa, T. (2008). Banana prevents plasma oxidative stress in healthy individuals. *Plant Foods for Human Nutrition*, 63: 71–76.
- Young, S. N. (2007). How to increase serotonin in the human brain without drugs. *Journal of Psychiatry & Neuroscience: JPN*, 32: 394–399.
- Zhang, P., Whistler, R. L., BeMiller, J. N., Hamaker, B. R. (2005). Banana starch: production, physicochemical properties, and digestibility—a review. *Carbohydrate Polymers*, 59: 443–458.