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PHYTOCHEMICAL STUDY AND ANTIBACTERIAL, ANTIFUNGAL AND ANTIOXIDANT PROPERTIES OF *BRIDELIA FERRUGINEA* AND *PTELEOPSIS SUBEROSA*

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ABSTRACT

The antifungal, antimicrobial, antioxidant and cytotoxic activities of methylene chloride, methanol and hydroethanolic extracts of the leaves of *Bridelia ferruginea* and stem bark of *Pteleopsis suberosa* were investigated against six Gram Positive and Gram negative strains bacteria and six species of *Aspergillus*. The antimicrobial activity was investigated by the microtest method using p-iodonitrotetrazolium and the antifungal activity by measuring the mycelial and sporulation inhibition. The phytochemical study was performed on Thin Layer Chromatography and antioxidant activity was assayed using qualitative and quantitative 1,1-diphenyl-2-picrylhydrazyl (DPPH) scavenging test. The methanolic extract of *Pteleopsis suberosa* showed better Minimum Inhibitory Concentration against all the tested bacteria. The strongest activity was observed with the methanolic extract of *Pteleopsis suberosa* against *S. aureus* *meticillin* resistant with Minimum Inhibitory Concentration value of 78µg/ml. All extracts showed an interesting sporulation inhibitory percentage up to 60% whereas little to moderate inhibition was obtained against mycelial development (9.18 to 57.53%). The antioxidant activity of extracts increased in dose dependent manner. The methanolic and hydroethanolic extracts were the most active with inhibitory percentage values varying from 86.09 to 100%. In the brine shrimp lethality bioassay, the LC₅₀ values of tested extracts ranged between 3.74 to 45.38 mg/ml. The results obtained indicated that the plant have interesting bioactive principles and support the use of these plants in the treatment of infectious diseases.

Keywords:

Antibacterial,
Antifungal,
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Pteleopsis. Suberosa,
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INTRODUCTION: The emergence of pathogens resistant to antibiotics represents a serious problem for public health. The resistance of pathogens such as bacteria and fungi to drugs is increasingly high. These pathogens caused major problems throughout the world. Unfortunately, a number of antibiotics produced by the pharmacological industries do not allow fighting the diseases caused by these pathogens.

Plants have been used for centuries as remedies for human diseases. As a result of increasing need for new and better antimicrobial and antifungal drugs, research work was carried out to study scientifically medicinal plants. Various antifungal and antimicrobial agents have been explored, but the control of fungal diseases has not yet been achieved¹⁻².

Single or combination of plants is used by traditional healers to treat various diseases. These traditional healers possess a good knowledge of the use of medicinal plants. They have also a number of superstitious beliefs related to different types of diseases and possess a good knowledge in using herbal drugs. Many Ethnobotanists all over the world stressed the importance of investigating plants for new antimicrobial and antifungal agents³⁻⁵.

The role of free radical reactions in disease pathology is also well established, suggesting that these reactions are necessary for normal metabolism but can be detrimental to health as well⁶. It is known that free radicals play a fundamental role in several diseases such as arteriosclerosis, cancer, diabetes mellitus, hypertension, renal failure, liver disease, AIDS etc⁷⁻¹⁰. Further to the remarkable toxicity and the mutagenic effects of synthetic antioxidants, the attention turned to the natural antioxidants. Numerous medicinal plants extracts or constituents have proven to show free radical scavenging activity^{5, 11-12}.

According to World Health Organization, 80% of the world population still relies mainly on plant drugs¹³.

Bridelia ferruginea (Euphorbiaceae) traditionally used by traditional healers is a shrub from 1 to 8 cm. The decoction of stem bark is used per os to treat epilepsy, oedemas, irritability of the infant. It was also used to treat gastralgias, anaemia, dysentery and rheumatism. The roots were also used with *Euphorbia hirta* to treat gonorrhoea and haemorrhage¹⁴.

Pteleopsis suberosa (Combretaceae) is a small tree used by traditional healers in Bénin. It is used traditionally to treat various diseases. The decoction of roots is used to treat dystocia and the stem bark to treat dysentery, eruptive fever, and epilepsy¹⁴.

The objective of the present study was to investigate the phytochemical constituents of *Pteleopsis suberosa* and *Bridelia ferruginea* and to evaluate the antibacterial, antifungal, antioxidant activities against seven bacteria strains, six species of *Aspergillus*.

The inhibitory effect of extracts against free DPPH radical and cytotoxicity on *Artemia salina* were also evaluated.

MATERIAL AND METHODS

Plant material: In the present study, the leaves of *Bridelia ferruginea* and bark of *Pteleopsis suberosa* were used. Plants were collected from Ouidah, in Atlantic Department, the southern commune of Bénin, in January 2010. The collection and identification were carried out by botanists from university of Abomey-Calavi. The parts collected for each plant were dried for two weeks in laboratory (22°C).

Preparation of Plants Extracts: The dried plants were ground to a fine powder using an electric grinder (EXCELLA Mixer Grinder with 3 S.S.JARS. Model: excella QTY:1PC). Air dried plant of each species (100 g) was extracted three times with methylene chloride (400 ml) at room temperature. The plant extracts were filtered through Whatman N°1 filter paper and pooled together. The filtrates were concentrated to dryness in a rotary evaporator at 40-50°C. The residue was dried under extractor hood and then extracted three times with methanol (400 ml) using same procedure. 50 g of each plant material were also extracted two times with 500 ml of 20% ethanol in water for 2 h. The filtrates of each extraction were taken to dryness under vacuum and the residues were stored in a freezer until used for biological assay.

Microorganisms: Seven bacteria strains: *Escherichia coli* CIP 53126, *Staphylococcus aureus* ATCC 6538, *Enterococcus faecalis* ATCC 29212, *Pseudomonas aeruginosa* CIP82118, *Salmonella aboni* CIP 8039, *Staphylococcus aureus* Meticillin Resistant (SARM), *Staphylococcus epidermidis* obtained from Laboratoire de Biophotonique et Pharmacologie, University of Strasbourg, France and Six fungal species, *Aspergillus flavus* CMBB75, *A. parasiticus* CMBB20, *A. ochraceus* CMBB91, *A. nidulans* CMBB90, *A. terreus* CMBB94 and *A. fumigatus* CMBB89 obtained from the Laboratory of Biochemistry and Molecular Biology of University of Abomey-calavi were used in the present study.

Phytochemical study: Phytochemical analysis for major constituents was achieved on Thin Layer Chromatography plate (Alugram, Silicagel 60 F₂₅₄) using a standard procedure¹⁵⁻¹⁶. The tests for alkaloids, coumarins, anthracenes derivatives, flavonoids, essential oils, lignans, naphthoquinones, pigments, saponins, terpenoids and tannins.

Minimum Inhibitory Concentration (MIC): The Minimum inhibitory concentrations were determined by serial dilution microplate bioassay using specific dye *p*-iodonitrotetrazolium violet as an indicator of growth¹⁷. All extracts were reconstituted to 20 mg/ml with a mixture of acetone/Muller Hinton (v/v). MIC was determined by two fold serial dilutions of extracts beyond the level where no inhibition of growth of microorganisms was observed. 100 µl of Muller Hinton (DIFCO: Becton Dickinson France S.A) broth culture of bacteria (10⁶ CFU/ml) was added to each well contains extract. The microplates were incubated at 37°C for 18 h, after which 40 µl of *p*-iodonitrotetrazolium (0.2 mg/ml) solution in water were added to each well and the microplates were incubated at 37°C. After 1 h of incubation, the MIC values were recorded. The total activities (TA) of extracts were determined by dividing the MICs with the quantity extracted from 1 g of the plant material¹⁸.

Antifungal Assay: Antifungal activity of extracts was evaluated against six species of *Aspergillus* as described by Saadabi¹⁹. The mixture of sterilized Potato Dextrose Agar-extract (PDA: 39 g of powder of PDA in 1 liter of distilled water) at 1 mg/ml was poured in sterile disposable petri dishes. After solidified, 100 spores prepared in tween 25% were dropping in the center on the petri dishes which were left at 25°C. After 5 days the diameter of mycelia was measured and the number of spores was counted microscopically using malassez cell. Each assay was run in triplicate. Two petri dishes without extract were used as control. The inhibitory percentage (*PI*) of extracts was determined according to the formula below²⁰:

$$PI (\%) = \frac{A - B}{A} \times 100$$

In which *PI* means: Inhibitory Percentage; *A* = Average diameter of the mycelia or estimated number of spores of control; *B* = Average diameter of the mycelia or estimated number of spores of tested dishes (with extract).

Qualitative Antioxidant Activity: The aim of this method was to evaluate the preliminary antioxidant activity before quantitative evaluation²¹. Each extracts were spotted on the starting point on silica gel sheet (Kieselgel 60 F₂₅₄, Merk) which was developed in the

mixture of Ethyl acetate/Methanol/water (100:2:1). After dry, the silica gel plate was sprayed with a solution of 2% DPPH in methanol. Any bleaching of the purple color background of DPPH (purple color) reagent within 10 min was taken as positive result.

Quantitative Antioxidant Activity: The quantitative antioxidant activity was determined according to the method described previously by Velazquez²². Three stock solutions of extracts (300; 30 and 3µg/ml) were prepared and tested in the final concentrations of 100, 10 and 1µg/ml. 750 µl of stock solution of each extract and 1500 µl of a 2% solution of DPPH in methanol were introduced into dry and sterile tubes. For each concentration a blank and a negative control are prepared. The Blank consists of 750 µl of extract and 1500 µl of methanol. The negative control consists of 1500 µl of the solution of DPPH (2%) and 750 µl of methanol. Each test was done in triplicate and quercetol was used as positive control. The test tubes were incubated in dark at room temperature. After 20 mn, the optical density of each mixture was measured at 517 nm using spectrophotometer (Jenway Jenova). The inhibitory Percentage of DPPH radical which means the antioxidant activity of extracts and quercetol was calculated as follow²³.

$$\%I = [1 - (DO_S - DO_B)/DO_C] \times 100$$

In which %I = Inhibitory Percentage of DPPH radical; DO_S = Absorbance of sample; DO_B = Absorbance of Blank; DO_C = Absorbance of control.

Brine Shrimp Lethality Bioassay: The assay was performed as described by Keymanesh *et al.* (2009). The brine shrimp eggs were hatched in normal seawater for 72 h to obtained nauplii larva. The stock solution of each extracts (3 mg/ml) was obtained by dissolving 15 mg in 200 µl ethanol and 4.80 ml of seawater. For the assay, 1 ml of seawater containing 15 living naupli to 1 ml of extracts at 3 mg/ml was added. Six concentrations ranging from 1.5 to 0.075 mg/ml; obtained by a twofold serial dilution of stock solution were tested. Each experiment was done in triplicate and control was prepared using seawater and 15 naupli. After 24 h incubation at room temperature, survivor's nauplii were counted. The Lethal concentration (LC₅₀) was determined graphically.

RESULTS AND DISCUSSION:

Phytochemical Results: Phytochemical analysis of *Bridelia ferruginea* and *Pteleopsis suberosa* are presented in **Table 1**.

TABLE 1: PHYTOCHEMICAL ANALYSIS OF BRIDELIA FERRUGINEA AND PTELEOPSIS SUBEROSA

	<i>Bridelia ferruginea</i>	<i>Pteleopsis suberosa</i>
Alkaloids	--	--
Coumarins	+	+
Anthracenic derivatives	++	--
Flavonoids	++	--
Essential oil	+++	++
Lignans	--	--
Naphtoquinones	++	--
Pigments	++	+++
Saponins	--	--
Triterpenoids	++	+++
Tannins	+	++

(--): Absence; (+): trace; (++): medium; (+++): presence

The phytochemical screening of *B. ferruginea* showed the presence of coumarins, anthracenic derivatives, flavonoids, essential oil, naphtoquinones, pigments, triterpens and tannins whereas *P. suberosa* showed the presence of coumarins, essential oil, pigments,

TABLE 2: MINIMUM INHIBITORY CONCENTRATION VALUES IN mg/ml AND TOTAL ACTIVITY IN ml OF EXTRACTS: DM: METHYLENE CHLORIDE, ME: METHANOL, ETOH/H₂O: ETHANOL/WATER

Species Extracts μorganismes	Minimum Inhibitory Concentration (mg/ml)					
	<i>Bridelia ferruginea</i>			<i>Pteleopsis suberosa</i>		
	DM	Me	EtOH/H ₂ O	DM	Me	EtOH/H ₂ O
<i>E.coli</i>	Nd	1.25	2,5	-	0.63	-
<i>S.aureus</i>	>2.5	1.25	2,5	-	0.15	-
<i>S.epidermidis</i>	>2.5	0.63	2.5	>2.5	0.31	0.63
<i>S.a Met Resistant</i>	1.25	0.31	0.63	0.16	0.078	0.16
<i>E.faecalis</i>	5	5	2.5	-	0.63	1.25
<i>P.aeruginosa</i>	5	2.5	2.5	-	1.25	-
<i>S.aboni</i>	2.5	5	-	-	2.5	0.63
Total quantity in mg extracted from 1 g						
	26.80	153.24	158.94	11.54	8.44	42.2
Total activity in ml/g						
<i>E.coli</i>	Nd	122.59	63.57	-	13.5	-
<i>S.aureus</i>	<10.72	122.59	63.57	-	54.01	-
<i>S.epidermidis</i>	<10.72	245.18	63,57	<4.61	27.01	67.52
<i>S.a Met Resistant</i>	21.44	490.36	254.3	73.85	108.20	270.08
<i>E.faecalis</i>	5.36	30.65	63.57	-	13.5	33.76
<i>P.aeruginosa</i>	5.36	61.29	63.57	-	6.75	-
<i>S.aboni</i>	10.72	30.65	-	-	3.37	67.52

triterpens and tannins. These results are comparable to those obtained by Owoseni and Baba-moussa respectively on ethanolic extract of leaves and barks of *Bridelia ferruginea* and *Pteleopsis suberosa*²⁴⁻²⁵.

Minimum Inhibitory Concentration (MIC): The Minimum Inhibitory Concentration (MIC) and Total Activity values are recorded in **Table 2**. The results showed that the MIC values ranging from 0.078 to 2.5 mg/ml. All extracts inhibited one or more microorganisms. The antibacterial activity of *P. suberosa* extracts was higher than *B. ferruginea* extracts. The methanolic extract of *P. suberosa* was the most active with MIC value of 78μg/ml against *Staphylococcus aureus* meticillin resistant (SARM) which was the most sensible among the tested bacteria while *E. faecalis* the most resistant. The results previously obtained with aqueous and ethanolic extracts of *B. ferruginea* are similar to those obtained in our study with the hydro-ethanolic extract against *E. coli*, *S. aureus* and *P. aeruginosa* (CMI > 470 μg/ml)²⁶⁻²⁷. The interesting antimicrobial activity obtained here could be due to the presence of flavonoids and tannins in the extracts²⁸⁻³⁰.

Antifungal Assay: The antifungal activity was evaluated against six *Aspergillus* species. Six extracts obtained from *Bridelia ferruginea* and *Pteleopsis suberosa* were tested for their effect against mycelial growth and sporulation of fungi. Results are compiled in **Table 3 and 4**. All tested extracts were active against fungi by inhibiting sporulation. The Inhibitory Percentage (IP) of

sporulation ranging from 53.98% to 99.09%. The most interesting activity was obtained with methanol extract of *B. ferruginea* against *Aspergillus flavus* with an IP value of 99.09%. *Aspergillus flavus* and *Aspergillus fumigatus* were the most sensitive for extracts (94.91% \geq IP \geq 99.09%).

Table 3: Antifungal activity of extracts on sporulation stage of *Aspergillus* species

Extracts	Inhibitory Percentage of sporulation (%)					
	<i>A. flavus</i>	<i>A. parasiticus</i>	<i>A. terreus</i>	<i>A. ochraceus</i>	<i>A. nidulans</i>	<i>A. fumigatus</i>
Bf DM	96,06 \pm 0,00	53,98 \pm 0,12	82,41 \pm 0,07	83,11 \pm 0,12	85,11 \pm 0,06	97,70 \pm 0,00
Bf Me	97,47 \pm 0,01	77,34 \pm 0,04	89,28 \pm 0,08	78,22 \pm 0,05	92,31 \pm 0,01	99,09 \pm 0,00
Bf H₂O	95,87 \pm 0,01	94,13 \pm 0,18	55,80 \pm 0,11	66,66 \pm 0,26	89,91 \pm 0,01	96,74 \pm 0,00
Ps DM	95,87 \pm 0,01	82,34 \pm 0,06	80,53 \pm 0,02	92,27 \pm 0,02	83,01 \pm 0,01	93,32 \pm 0,00
Ps Me	96,92 \pm 0,01	89,69 \pm 0,03	80,53 \pm 0,01	86,30 \pm 0,07	86,37 \pm 0,03	70,14 \pm 0,01
Ps H₂O	94,91 \pm 0,01	62,01 \pm 0,12	86,16 \pm 0,02	86,30 \pm 0,04	58,05 \pm 0,04	69,57 \pm 0,06

(Bf): *Bridelia ferruginea*; (Ps): *Pteleopsis suberosa*; (DM): Dichlorométhane, (Me): Méthanol; EtOH/H₂O: Ethanol/Eau

TABLE 4: ANTIFUNGAL ACTIVITY OF EXTRACTS AGAINST MYCELIAL DEVELOPMENT STAGE OF ASPERGILLUS SPECIES.

Extracts	Inhibitory Percentage of mycelia development (%)					
	<i>A. flavus</i>	<i>A. parasiticus</i>	<i>A. terreus</i>	<i>A. ochraceus</i>	<i>A. nidulans</i>	<i>A. fumigatus</i>
<i>Bf</i> DM	18,36 \pm 0,1	27 \pm 0,01	41,1 \pm 0,06	34,48 \pm 0,08	7,46 \pm 0,02	11,36 \pm 0,22
<i>Bf</i> Me	20,41 \pm 0,0	41 \pm 0,12	46,57 \pm 0,0	48,27 \pm 0,02	32,83 \pm 0,03	34,10 \pm 0,10
<i>Bf</i> H ₂ O	12,24 \pm 0,0	27 \pm 0,09	57,53 \pm 0,0	43,1 \pm 0,00	43,28 \pm 0,15	36,36 \pm 0,13
<i>Ps</i> DM	24,44 \pm 0,0	29 \pm 0,01	46,57 \pm 0,0	50,00 \pm 0,04	35,82 \pm 0,03	11,36 \pm 0,14
<i>Ps</i> Me	19,38 \pm 0,1	28 \pm 0,02	45,2 \pm 0,01	43,10 \pm 0,05	31,34 \pm 0,09	27,27 \pm 0,22
<i>Ps</i> H ₂ O	9,18 \pm 0,01	27 \pm 0,04	36,98 \pm 0,0	39,65 \pm 0,05	37,31 \pm 0,01	15,90 \pm 0,21

(Bf): *Bridelia ferruginea*; (Ps): *Pteleopsis suberosa*; (DM): Dichlorométhane ; (Me): Méthanol; (EtOH/H₂O): Ethanol/Eau

Contrary to sporulation, the Inhibitory Percentage (IP) of extracts against mycelial development was weak (2.98% to 53.42%). Only 11 tests out of 36 showed an IP value up to 40%. The hydroethanolic extract of *B. ferruginea* and methylene chloride extract of *P. suberosa* showed interesting activity with IP values of 57.53% and 50.00% respectively. The other extracts showed weak results (7.46% - 48.27%).

Antioxidant activity: Antioxidant activity of extracts was determined for their scavenging potential of the stable DPPH free radical in both qualitative and quantitative tests.

- **Qualitative antioxidant activity of extracts:** Figure 1 showed the qualitative inhibition of DPPH radical by extracts. The results obtained showed that methanolic and hydroethanolic extracts of both species presented the most interesting activities. Methylene chloride extracts showed respectively weak to low inhibition of DPPH radical. In the works of Dramane, the methanolic extract of bark of *B. ferruginea* gave similar positive results²⁷.



FIGURE 1: QUALITATIVE ANTIOXIDANT EFFECT OF EXTRACTS FROM B. FERRUGINEA AND P. SUBEROSA

- **Quantitative Antioxidant Activity:** Figure 2 shows the antioxidant potential of six extracts from *B. ferruginea* and *P. suberosa* expressed as Inhibitory percentage of DPPH radical in comparison to the positive control (quercetol). The results obtained showed that the antioxidant activity of extracts is concentration dependent. It has been shown that the scavenging effects on the DPPH radical increase with increasing concentration of samples and standards to a certain extent²⁸.

The methanol and hydroethanol extracts of both species showed the most interesting antioxidant activity. At 1µg/ml, the antioxidant activity turns around 10% whereas quercetol showed an inhibitory percentage of 74.5%. Methylene chloride extract of *B. ferruginea* possesses no activity which result was in accordance with that obtained by the qualitative method. At 10µg/ml, four extracts out of six presented an antioxidant activity up to 40%. The hydroethanolic extract was the most active with an IP value of 91.89%.

At 100 µg/ml, except methylene chloride extract of *B. ferruginea* (20, 79%), the other five extracts showed inhibitory percentages ranging from 86 to 100%, similar to quercetol (86%). Our results suggested that *B. ferruginea* and *P. suberosa* contain compounds with strong radical scavenging activity.

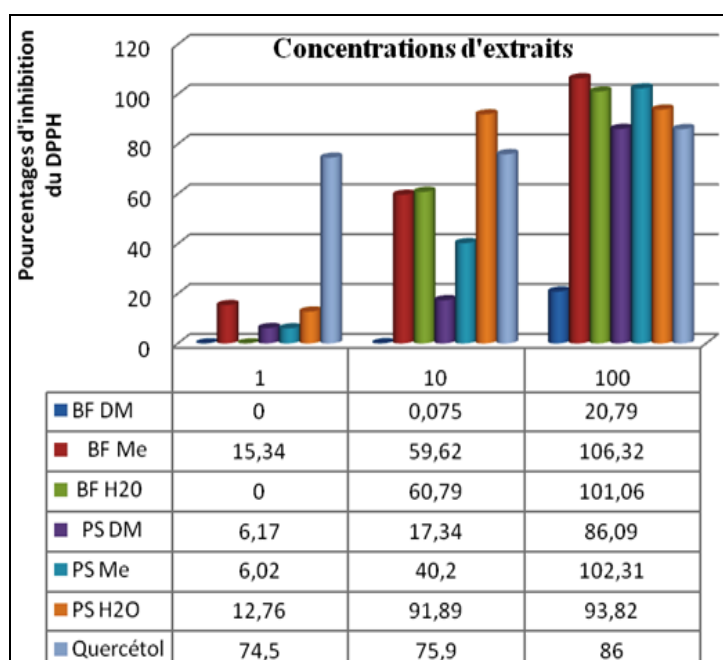


FIGURE 2: QUANTITATIVE ANTIOXIDANT EFFECT OF EXTRACTS FROM *B. FERRUGINEA* AND *P. SUBEROSA*

Brine Shrimp Lethality Bioassay: Brine shrimp lethality test results are showed in Table 5. The LC₅₀ values of tested extracts ranged between 3.74 to 45.38 mg/ml. The hydroethanolic extract was the most toxic on the shrimps with LC₅₀ value of 3.74 mg/ml. The methylene chloride extracts of both species showed similarly LC₅₀ values of 22.37 and 20.42 mg/ml respectively for *B. ferruginea* and *P. suberosa*. The higher LC₅₀ value was exhibited by methanol extract of *P. suberosa* (45.38 mg/ml). According to the results previously obtained by Zakaria²⁹ we concluded that all extracts tested in his study exhibited no toxicity, giving LC₅₀ values higher than 100µg/ml.

TABLE 5: BRINE SHRIMP LETHALITY ASSAY OF EXTRACTS FROM *B. FERRUGINEA* AND *P. SUBEROSA*

Species	Extracts	LC ₅₀	Coefficient co-relation
<i>Bridelia ferruginea</i>	DM	22,37	0,68
	Me	13,63	0,58
	EtOH/H ₂ O	3,74	0,88
<i>Pteleopsis suberosa</i>	DM	20.42	0,68
	Me	45,38	0,38
	EtOH/H ₂ O	Nt	0

CONCLUSION: In the present study, biological activity of six extracts from *B. ferruginea* and *P. suberosa* was investigated. We can conclude that both species may have antibiotic and antifungal activity as they exhibited potent antibacterial, antifungal, antioxidant activities. Chance to find constituents with antifungal, antimicrobial activities were apparent in the two species. These findings give a scientific basis to the traditional uses of *B. ferruginea* and *P. suberosa*.

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REFERENCE:

1. Goun E, Cunningham G, Chu D, Nguyen C and Miles D: Antibacterial and antifungal activity of Indonesian ethnomedical plants. *Fitoterapia* 2003; 74: 592-596.
2. Omer MEF and Elnima EI: Antimicrobial activity of *Ximenia Americana*. *Fitoterapia* 2003; 74: 122-126.
3. Adams M, Gmunder F and Hamburger M: Plants traditionally used in age related brain disorders-A survey of ethnobotanical literature. *J. Ethnopharmacol* 2007; 113: 363-381.
4. Sri Rama MK and Nagamani K: Antimicrobial Spectrum and Phytochemical Study of *Walsura trifoliata* (A. Juss) Harms.

- (Meliaceae) Bark Extracts. Journal of Pharmacology and Toxicology 2008; 3 (4): 267-271.
5. Lagnika L, Anago E, Atindehou M, Adjahoutonon B, Dramane K and Sanni A: Antimicrobial activity of *Crataeva religiosa* Forst against bacteria isolated from *Thryonomys swinderianus* Temminck. African Journal of Biotechnology 2011; 10(49): 10034-10039.
 6. Gupta M, Mazumder UK and Gomathi P: Evaluation of antioxidant and free radical scavenging activities of *Plumeria acuminata* leaves. Journal of Biological Sciences 2007; 7 (8): 1361-1367.
 7. Govindarajan R, Vijayakumar M and Pushpangadan P: Antioxidant approach to disease management and the role of rasayana herbs of Ayurveda. J. Ethnopharmacology 2005; 99: 165-178.
 8. Tiwari AK: Antioxidants: New-generation therapeutic base for treatment of polygenic disorders. Current Science 2004; 86: 1092-1102.
 9. Cheng HY, Lin TC, Yu KH, Yang CM and Lin CC: Antioxidant and free radical scavenging activities of *Terminalia chebula*. Biol. Pharm. Bull 2003 ; 26 : 1331-1335.
 10. Ellnain-Wojtaszek M, Kruczynski Z and Kasprzak J: Investigation of the free radical scavenging activity of *Ginkgo Biloba* L. leaves. Fitoterapia 2003; 74: 1-6.
 11. Rahman I: Antioxidant therapeutic advances in COPD. Ther. Adv. Respir. Dis 2008; 2: 351 - 374.
 12. Trichopoulou A, Soukara S and Vasilopoulou E: Traditional foods: A science and society perspective. Trends Food Sci. technol. 2007 ; 18: 420-427.
 13. Kumara PD, Jayawardane GL and Aluwihare AP: Complete colonic duplication in an infant. Ceylon Med. J. 2001; 46: 69-70.
 14. Adjanohoun EJ, Adjakidjè V, Ahyi MRA, Akeassi L, Akoegninou A, d'Almeida J, et al: Médecine traditionnelle et pharmacopée. Contribution aux études ethnobotaniques et floristiques en République du Bénin. Paris, Agence de Coopération Culturelle et Technique, pp. 339. 1989.
 15. Wagner H and Blat S: Plant Drug Analysis. Springer, 2nd edition 2001.
 16. Bruneton J: Pharmacognosie, Phytochimie, Plantes médicinales. TEC DOC Paris France, 4th edition 2009.
 17. Eloff JN: A sensitive and quick method to determine the minimal inhibitory concentration of plant extracts for bacteria. Planta Medica 1998; 64: 711-713.
 18. Eloff JN: Quantification the bioactivity of plant extracts during screening and bioassay guided fractionation. Phytomedicine 2004; 11: 370-371.
 19. Saadabi AMA: Antifungal activity of some Saudi plants used in traditional medicine, Asian Journal of Plant Science 2006; 5 (5), 907-909.
 20. Dohou N, Yamni K, Badoc A and Douira A : activité antifongique d'extraits de *Thymelaea lythroides* sur Trois champignons pathogènes du riz. Bull. Soc. Pharm. 2004 ; 143 : 31-38.
 21. Rached W, Benamar H, Bennaceur M and Marouf A: Screening of the antioxidant potentiel of some algerian indigenous plants. Journal of Biological Sciences 2010; 10 (4): 316-324.
 22. Velazquez E, Tournier HA, Mordujovich de Bushiazzo P, Saavedra G and Schinella GR: Antioxidant activity of Paraguayan plants extracts. Fitoterapia 2003; 74: 91-97.
 23. Schmeda-Hirschmann G, Rodriguez J, Theoduloz C, Astudillo S, Feresin G and Tapia A: Free-radical scavengers and antioxidants from *Peumus boldus* Mol (Boldo). Free Radical Res. 2003; 37: 447-452.
 24. Owoseni AA, Ayanbamiji TA, Ajayi YO and Ikeoluwa B: Antimicrobial and phytochemical analysis of leaves and bark extracts from *Bridelia ferruginea*. African Journal of Biotechnology 2010; 9 (7): 1031-1036.
 25. Baba-Moussa F Akpagana K and Bouchet P: Antifungal activities of seven West African Combretaceae used in traditional medicine. Journal of Ethnopharmacology 1999; 66 (3): 335-338.
 26. Adewale A, Winston AM and Corcoran O: Antibacterial, antioxidant and fibroblast growth stimulation activity of crude extracts of *Bridelia ferruginea* leaf, a wound-healing plant of Nigeria. Journal of Ethnopharmacology 2011; 133(1): 116-119.
 27. Dramane S, Witabouna KM and Kamanzi K: Evaluation des Activités Antimicrobiennes et Anti-Radicaux Libres de Quelques Taxons Bioactifs de Côte D'ivoire. Journal of Scientific Research 2010 ; 40 (0): 307-317.
 28. Motalleb G, Hanachi P, Kua SH, Fauziah O and Asmah R: Evaluation of phenolic content and total antioxidant activity in *Berberis vulgaris* fruits extract. J. Biol. Sci. 2005; 5: 648-653.
 29. Zakaria HM, Mainen JM, Pax JM, Modest CK and Ramadhani SON: Antimicrobial activity and brine shrimp toxicity of extracts of *Terminalia brownii* roots and stem. BMC Compl. Altern. Med. 2007; 7: 9.

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